ORIGINAL ARTICLE

Forensic and phylogeographic characterization of mtDNA lineages from northern Thailand (Chiang Mai)

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Abstract The immigration of diverse ethnic groups over the past centuries from surrounding countries into Thailand left footprints in the genetic composition of Thai mitochondrial DNA (mtDNA) lineages. The entire mtDNA control region (1,122 bp) was typed in 190 unrelated male volunteers from the northern Thailand province of Chiang Mai following highest quality standards. For a more precise haplogroup classification, selected single nucleotide polymorphisms from the mtDNA coding region were genotyped. We found several new, so far undescribed mtDNA lineages. Quasi-median networks were constructed for visualisation of character conflicts. The data were put into population-genetic relationships with other Southeast Asian populations. Although the frequencies of the Thai haplogroups were characteristic for Southeast Asia in terms of haplotype

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composition and genetic structure, the Thai population was significantly different from other Southeast Asian populations. This necessitates establishing regional databases, especially for forensic applications. The population data have been submitted to the EMPOP database (www.empop.org) and will be available on publication.

Keywords mtDNA population data · Southeast Asia · Network analysis · Haplogroup assignment · Phylogeographic analyses · Forensic science

Introduction

The Kingdom of Thailand is located in the heart of Southeast Asia. Due to its geographical location, Thai culture has always been under strong influence from China and India. Thailand is the only Southeast Asian country

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never colonised by a European power [1]. Thailand's population is dominated by various peoples speaking Tai-Kadai, which is one of the major language families of Southeast Asia [2]. Other resident ethnic groups in Thailand include Malays, Mon, Khmer, and various hill tribes [3].

The Chinese colonisation of Thailand started during the early second millennium AD when people migrated from southern China into Southeast Asia. During the twelfth and thirteenth centuries, Thai people were subjugated under the Khmers leading to an evident mitochondrial DNA (mtDNA) exchange between the Tai-Kadai and the Khmer groups [4]. In addition, Thailand has a long tradition of granting political asylum to religious or ethnically hunted refugees from neighbouring countries. Vietnamese Christians, Mon people from Burma, and political dissidents from Cambodia have sought and received shelter in Thailand since hundreds of years [5]. The genetic diversity of northern Thailand hill tribes, in particular, was additionally affected by sex-specific migration rates and cultural factors [3].

The consequences of this ethnic amalgamation on the genetic structure of Thailand are widely unknown. Available mitochondrial genetic data of Thailand's population is scarce, restricted by small sample sizes and limited to mitochondrial regions HVS-I [3, 4, 6–8] and HVS-I and HVS-II [9–11]. In this population study, 190 unrelated individuals living in the province of Chiang Mai (Fig. 1) were analysed, and their genetic structure was put into a population-genetic context with surrounding Asian populations.

Materials and methods

Samples, DNA extraction, and control region sequence analysis

Blood samples were taken from 190 unrelated male volunteers from small rural villages throughout the northern Thai province of Chiang Mai. DNA was isolated from peripheral blood lymphocytes according to standard procedures [12]. The entire mtDNA control region (CR; nps 16024–16569; 1–576) was amplified, sequenced, and evaluated following EMPOP recommendations [13, 14] and updated nomenclature guidelines for mtDNA [15].

Haplogroup assignment and analysis of coding region SNPs

MtDNA CR haplotypes were affiliated to haplogroups based on patterns of shared haplogroup-specific or -associated polymorphisms [16–23]. To confirm and specify the haplogroup assignment derived from the CR patterns, we analysed selected coding region single nucle-



Fig. 1 Map of Thailand. Note: The province of Chiang Mai is *shaded in grey*

otide polymorphisms (SNPs) targeting haplogroup-specific mutations in Southeast Asian lineages using a modified version of the SNaPshot assay presented in [24]. Furthermore, we developed a sequencing-based screening method for haplogroup M, N, and C-specific coding region SNPs. The analysed SNPs were selected on the basis of published phylogenetic trees [16–23, 25].

In paragroup N* samples, we examined two coding region fragments ranging from nps 4450-5860 and from nps 6340-7770. For unresolved or ambiguous paragroup M* associated samples, we determined the sequences of

Table 1 Diversity measures from various ethnic populations from Thailand

| Population statistics | Chiang Mai | Akha | Chiang Rai Lisu | Black Lahu | Mae Hong Son Lisu | Red Karen | White Karen | Various ethnic groups of Thailand |
|--------------------------------|-------------|---------------|--------------------|---------------|----------------------|---------------|---------------|---|
| N | 190 | 91 | 53 | 39 | 42 | 39 | 40 | 215 |
| Reference | This study | Oota 2001 [6] | Oota 2001 [6] | Oota 2001 [6] | Oota 2001 [6] | Oota 2001 [6] | Oota 2001 [6] | Fucharoen 2001 [9] |
| Number of haplotypes | 124 | 28 | 28 | 11 | 19 | 14 | 17 | 131 |
| Number of unique haplotypes | 95 | 12 | 20 | 6 | 7 | 8 | 10 | 125 |
| Haplotype diversity | 0.985 | 0.928 | 0.909 | 0.834 | 0.932 | 0.839 | 0.891 | 0.992 |
| Mean pairwise differences | 7.498±3.518 | 5.616±2.720 | 6.307±3.039 | 6.0256±2.934 | 5.843±2.849 | 4.626±2.319 | 5.108±2.529 | 7.016±3.302 |

Analysed range, 16048-16365

four fragments within the coding region (nps 1820–2450, nps 4450-5580, nps 8670–9850, and nps 11450–12820). Furthermore, one fragment was investigated ranging from nps 5820–6660 for samples assigned to lineage C*.

These target fragments were amplified in singleplex polymerase chain reaction (PCR) reactions in a total volume of 25 μ l under following conditions: 95°C for 2 min followed by 35 cycles of 95°C for 15 s, 56°C for 30 s, and 72°C for 90 s, followed by an extension phase at 72°C for 10 min. Purification of PCR products, cycle sequencing, and purification of cycle sequencing products were carried out following standard procedures [14]. The amplification and sequencing primer sequences applied are listed in Table S1a and S1b, respectively.

Random match probability

The random match probability was calculated as the sum of squared haplotype frequencies [26] based on mtDNA CR sequences (C-insertions in length-heteroplasmic regions around nps 16193, 309, and 573 were disregarded).

Population-genetic analyses

Molecular diversity indices, pairwise differences between and within populations, and an analysis of molecular

Table 2 Diversity measures from Southeast Asian populations

variance (AMOVA) were calculated using ARLEQUIN (version 3.11). The Chiang Mai samples from northern Thailand were compared with hill tribes and other ethnic groups of Thailand (Table 1). All sequences were aligned and trimmed to a greatest common range of nps 16048–16365, and length-heteroplasmic C-insertions around np 16193 were disregarded.

In addition, the Thailand samples were compared with surrounding Southeast Asian populations (Table 2). All sequences were aligned and trimmed to a greatest common range of nps 16024–16365 and nps 73–340; length-heteroplasmic C-insertions around nps 16193 and 309 were disregarded.

Results and discussion

Haplogroup structure and most common haplotypes

Within a total of 190 samples from Chiang Mai, 145 distinct CR haplotypes (disregarding C-insertions around positions 16193, 309, and 573) were found (Table S2). The proportions of the macrohaplogroups M, N, and R were 39.5%, 7.9%, and 52.6%, respectively. The most common haplotype was F1a (9.5%; 16129A-16172C-16304C-16519C-73G-249DEL-263G-315.1C-523DEL-524DEL),

| Population statistics | Thailand | Japan | N-China | Korea | Taiwan | Vietnam | Malaysia |
|------------------------------|--------------|--------------------|----------------|---------------|-------------------|-----------------|----------------|
| N | 190 | 162 | 232 | 593 | 640 | 187 | 205 |
| Reference | This study | Imaizumi 2002 [27] | Kong 2003 [28] | Lee 2006 [29] | Trejaut 2005 [30] | Irwin 2008 [31] | Wong 2007 [32] |
| Number of haplotypes | 137 | 131 | 129 | 408 | 339 | 154 | 152 |
| Number of unique haplotypes | 108 | 119 | 80 | 325 | 231 | 136 | 115 |
| Haplotype diversity | 0.989 | 0.988 | 0.988 | 0.995 | 0.994 | 0.991 | 0.991 |
| Mean pairwise differences | 10.213±4.684 | 8.878±4.114 | 8.427±3.914 | 9.594±4.405 | 9.790±4.489 | 10.366±4.750 | 10.680±4.883 |

Analysed range, 16024-16365 and 73-340

followed by F1a1a (7.9%; 16108T-16129A-16162G-16172C-16304C-16519C-73G-249DEL-263G-315. 1C-523DEL-524DEL) and M7b1 (6.8%; 16129A-16192T-16223T-16297C-73G-150T-199C-263G-315.1C-489C).

Haplogroups C, G, A, and Z each occurred at frequencies lower than 3%. We found five remarkable lineages within macrohaplogroups M and N that were new to the current literature; until their phylogenetic position is clarified, we denote those lineages as "macrohaplogrouppolymorphic position" (e.g., "N-5120"). A detailed list of all haplogroups and their frequencies are given in Table 3 and Fig. 2, respectively.

Effectiveness of targeted coding region sequencing

All samples were sequenced in the entire CR and SNPgenotyped with an extended version of the multiplex assay presented in [24]. For the majority of samples (72.6%), the haplogroup information from the SNP multiplex genotyping assay confirmed the haplogroup status of CR sequencing but did not lead to a higher resolution. For 12.6% of samples, however, the SNP multiplex provided a more precise haplogroup affiliation. In the majority of remaining samples (14.8%), targeted coding region sequencing enabled a more detailed haplogroup assignment.

Random match probability and mean number of pairwise differences

The estimated probability of a random match between two unrelated individuals from the Chiang Mai dataset was 1.04%, corresponding to a power of discrimination of 98.96% for the entire CR, when ignoring length variants around positions 16193, 309, and 573. The mean number of pairwise differences for this dataset was calculated 11.56 ± 5.26 ignoring hotspot insertions.

Comparison with other ethnic groups from Thailand

To shed more light on the genetic composition in Thailand, we compared the current sample set from Chiang Mai, northern Thailand, with six distinct ethnic groups and a compiled sample set of various ethnic groups of Thailand (Table 1). The highest intra-population diversity was found within our sample set from Chiang Mai with an average number of pairwise differences around 7.5 followed by the sample set from [9] (7.0). The lowest intra-population diversity was found in the matrilocal group of Red Karen with an average number of pairwise differences around 4.6. Twenty-seven haplotypes (21.8%) of Chiang Mai were found in other populations from Thailand (Table S3a). All six ethnic groups (Akha, Lisu from Chiang Rai and from Mae Hong Son, Black Lahu, Red Karen, and White Karen)

Table 3 Haplogroup frequencies of 190 samples from northernThailand (Chiang Mai)

| Haplogroup | Ν | Frequency (%) |
|-------------|----|---------------|
| A | 3 | 1.6 |
| B* | 1 | 0.5 |
| B4* | 1 | 0.5 |
| B4a | 4 | 2.1 |
| B4b | 4 | 2.1 |
| B4c | 4 | 2.1 |
| B4d | 4 | 2.1 |
| B4g | 2 | 1.1 |
| B5a | 10 | 5.3 |
| C7 | 5 | 2.6 |
| D4 | 8 | 4.2 |
| D5 | 2 | 1.1 |
| F* | 2 | 1.1 |
| F1* | 1 | 0.5 |
| F1a* | 18 | 9.5 |
| F1a1* | 1 | 0.5 |
| Flala | 15 | 7.9 |
| Flalc | 1 | 0.5 |
| F1a2 | 3 | 1.6 |
| F1b′d | 1 | 0.5 |
| F1c | 1 | 0.5 |
| F2 | 4 | 2.1 |
| F3a | 8 | 4.2 |
| G2 | 5 | 2.6 |
| M* | 8 | 4.2 |
| M14 | 1 | 0.5 |
| M-5054 | 4 | 2.1 |
| M7b* | 8 | 4.2 |
| M7b1 | 13 | 6.8 |
| M7b'd | 1 | 0.5 |
| M7c | 2 | 1.1 |
| M-8718* | 4 | 2.1 |
| M-8718-9548 | 8 | 4.2 |
| M9 | 5 | 2.6 |
| N* | 1 | 0.5 |
| N-5120 | 1 | 0.5 |
| N-5581 | 5 | 2.6 |
| N9a | 3 | 1.6 |
| pre-N22 | 2 | 1.1 |
| R* | 1 | 0.5 |
| R11 | 1 | 0.5 |
| R2 | 2 | 1.1 |
| R22 | 1 | 0.5 |
| R9 | 10 | 5.3 |
| Z | 1 | 0.5 |

Fig. 2 Schematic tree of mitochondrial lineages in a population sample from Chiang Mai, Thailand (*N*=190). Note: The size of the spheres portrayed corresponds to haplogroup frequencies. *Dark grey*: descendants from haplogroup M; *light grey*: descendants from haplogroup R; *white*: descendants from haplogroup N. The tree is rooted in L3



and the population compilation from [9] share various haplotypes at similar proportions with the sample from Chiang Mai, thus affirming its representativeness for northern Thai populations. AMOVA was used to test for significant variation in the mtDNA genetic structure among the various Thai populations (Table S4). Most of the observed genetic variation was attributable to differences within populations (95.0%). Variance among populations accounted for 5.0% (Table S4a). Although all of these populations were from the same geographical region, they differed highly significantly in their genetic structure (Table S4b), which can be explained by their population history (agricultural versus partly nomadic life style) and different social structures. The population average pairwise differences (Table S4c) showed that the mean number of pairwise differences for the Chiang Mai sample was lowest with the White Karen population. However, when using corrected values, the observed and the expected number of pairwise differences was closest between Chiang Mai and the composition of various ethnic groups from [9], thus further demonstrating the similarity in genetic structure of these two complex random population samples.

Comparison with other Southeast Asian populations

We found 19 shared haplotypes (approximately 14%) in the Thai and other Southeast Asian populations (Table S3b). AMOVA was used to test for significant variation in the mtDNA distributions among the various Southeast Asian populations (Table S5). The vast majority of the observed variance (96.3%) within these six populations was attributable to differences within populations, and 3.7% represented differences among populations (Table S5a).

One of the most common haplotypes (comparison range, 16048–16365), a haplogroup F1a lineage (16129A-16172C-16304C-16519C) found in the population from Chiang Mai was also found in several Thai groups (Akha, Black Lahu, Red Karen, White Karen, and in the data set from [9]), as well as in Vietnam and Malaysia. Also, one haplogroup M7b1 lineage (16129A-16192T-16223T-16297C) was found several times in the dataset from [9] and in the datasets from northern China and Vietnam. Interestingly, haplogroup F1a1a, clearly defined by CR polymorphisms 16108T and 16162G, could only be found in the Thai populations and not in the surrounding Asian

populations. A haplotype that only occurred once in our Chiang Mai sample (16223T-16311C-16362C-16519C) seemed to be common for Southeast Asia since it was found in Malaysia (twice) and in the distant Korean population (once), as well as in the group of Akha (observed 12 times), in the White Karen (observed five times), and in the compilation of [9] (observed four times; see Tables S3b).

Conclusions

The mtDNA CR sequences presented in this study were generated according to high-quality laboratory standards. The frequencies of the different haplogroups were characteristic for Southeast Asian populations. However, the Thai population was significantly different both in terms of haplotype composition and genetic structure. This necessitates establishing regional databases especially for forensic database searches in order to get reliable frequency estimates. From a phylogenetic point of view, it was interesting to find new, so far undescribed lineages. More sequences from such haplotypes need to be collected and typed for the entire mtDNA genome in order to expand the Southeast Asian mtDNA tree. The haplotypes reported in the present study will be made available from the EMPOP database (www.empop.org) on publication.

This publication follows the recommendations of the International Society for Forensic Genetics on the use of mtDNA in forensic analysis.

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